IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Pat nt Examining Operations

Application of:	Johnson		
Serial No:	09/158,120	Art Unit:	1644
Filed:	September 21, 1998	Examiner:	Roark
Title:	Human-Murine Chimeric Antibodies Against Respiratory Syncytial Virus		
Attorney Docket No.:	469201-367	Customer No.	27162

#22/E

TRANSMITTAL LETTER

Commissioner for Patents
Washington, D.C. 20231

SIR:

Enclosed please find the following:

1. Clean and Marked-up Copies of Page 17, as Amended; and
2. A self-addressed, postage paid, return receipt postcard, date stamp and return of which is respectfully requested.

The Commissioner is authorized to charge payment of any additional filing fees required under 37 C.F.R. 1.16 associated with this communication or credit any overpayment to Deposit **Account No. 03-0678**.

FIRST CLASS CERTIFICATE

I hereby certify that this correspondence is being deposited today with the U.S. Postal Service as First Class Mail in an envelope addressed to:

Commissioner for Patents
Washington, D.C. 20231

Raymond J. Lillie 10/15/02
Raymond J. Lillie, Esq. Date

Respectfully submitted,

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#157386 v1

SJ154
GGCGTCGACTCACCATGGACATGAGGGTCC(C/T)CGCTCAGC

SJ155 (H1129L CDR 1)
GTCACCATCACTTGCAAGTGCCAGCTGAGTGTAGGTTACATGCACTGGTACC
AGCAG (SEQ ID NO:10)

SJ157 (H1129L CDR 3)
GCAACTTATTACTGCTTTCAGGGGAGTGGGTACCCATTCACGTTCGGAGGGG
GG (SEQ ID NO:11)

SJ168
GTGACCAACATGGACCCTGCTGATACTGCCAC (SEQ ID NO:12)

SJ169
CCATGTTGGTCACTTTAAGGACCACCTGG (SEQ ID NO:13)

SJ170
CCAGTTTACTAGTGTCATAGATCAGGAGCTTAGGGGC (SEQ ID NO:14)

SJ171
TGACACTAGTAAACTGGCTTCTGGGGTCCCATCAAGG (SEQ ID NO:15)

PCR conditions

0.5uL of 1st strand cDNA, 10mM Tris-HCl pH8.3, 50mM KCl, 1.5mM Mg2Cl, 0.2mM dNTP's, 0.001 % gelatin, 1 uM each primer, 1 ng DNA template and 2.5u AmpliTaq(TM) DNA polymerase (Perkin Elmer - Cetus). 94° 1 minute, 55° 2 minutes, 72° 2 minutes in Perkin Elmer 480 thermocycler for 25 cycles. The resulting DNA fragment(s) were then extracted once with phenol/chloroform (1/1), precipitated with 2.5 volumes of ETOH, resuspended in the appropriate restriction endonuclease buffer and digested with restriction endonucleases to produce cohesive ends for cloning. The resulting fragments were then separated by electrophoresis on a 1 % agarose gel. After staining the gel with ethidium bromide the fragments were excised and purified from the agarose by freezing and extraction in the presence of phenol.

The fragments were then digested with restriction endonucleases EcoRI and BamHI and cloned into plasmid pUC18. The inserts were

SJ154
GGCGTCGACTCACCATGGACATGAGGGTCC(C/T)CGCTCAGC

SJ155 (H1129L CDR 1)
GTCACCATCACTTGCAAGTGCCAGCTGAGTGTAGGTTACATGCACTGGTACC
AGCAG (SEQ ID NO:10)

SJ157 (H1129L CDR 3)
GCAACTTATTACTGCTTTCAGGGGAGTGGGTACCCATTCACGTTCGGAGGGG
GG (SEQ ID NO:11)

SJ168
GTGACCAACATGGACCCTGCTGATACTGCCAC (SEQ ID NO:12)

SJ169
CCATGTTGGTCACTTTAAGGACCACCTGG (SEQ ID NO:13)

SJ170
CCAGTTTACTAGTGTCATAGATCAGGAGCTTAGGGGC (SEQ ID NO:14)

SJ171
TGACACTAGTAAACTGGCTTCTGGGGTCCCATCAAGG (SEQ ID NO:15)

PCR conditions

0.5uL of 1st strand cDNA, 10mM Tris-HCl pH8.3, 50mM KCl, 1.5mM Mg2Cl, 0.2mM dNTP's, 0.001 % gelatin, 1 uM each primer, 1 ng DNA template and 2.5u AmpliTaq(TM) DNA polymerase (Perkin Elmer - Cetus). 94° 1 minute, 55° 2 minutes, 72° 2 minutes in Perkin Elmer 480 thermocycler for 25 cycles. The resulting DNA fragment(s) were then extracted once with phenol/chloroform (1/1), precipitated with 2.5 volumes of ETOH, resuspended in the appropriate restriction endonuclease buffer and digested with restriction endonucleases to produce cohesive ends for cloning. The resulting fragments were then separated by electrophoresis on a 1 % agarose gel. After staining the gel with ethidium bromide the fragments were excised and purified from the agarose by freezing and extraction in the presence of phenol.

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